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Multistalides A and B, two novel sesquiterpenoid dimers from *Chloranthus multistachys*

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ABSTRACT

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Two novel sesquiterpenoid dimers, multistalides A and B (1–2), were isolated from the whole plant of *Chloranthus multistachys*. Their structures were established on the basis of spectroscopic analysis, and the absolute configuration of 1–2 was determined by CD exciton chirality method. © 2009 Elsevier Ltd. All rights reserved.

The genus *Chloranthus* (Chloranthaceae) is mainly distributed in eastern Asia,¹ and there are 13 species and five varieties found in China.² Most of these plants have been used as a folk medicine for the treatment of bone fracture.³ Previous chemical investigations on this plant genus have resulted in the isolation of an array of sesquiterpenoids,⁴ sesquiterpenoid oligomers,⁵ and diterpenoids.⁶ Recently, several studies showed that the sesquiterpenoid dimers isolated from the genus *Chloranthus* exhibited significant bioactivities.^{5e-i}

Chloranthus multistachys Pei, a major species of the genus *Chloranthus*, grows favorably in the damp areas of eastern Asia.¹ Our preliminary study on this plant with a small amount of material has led to the isolation of one new sesquiterpenoid dimer^{5b} and several diterpenoids.^{6b} In a continuing phytochemical study, the whole plants of *C. multistachys* were re-examined, two new sesquiterpenoid dimers, multistalides A and B (**1–2**) were isolated. Their structures were established on the basis of spectroscopic analysis, and their absolute stereochemistry was determined by CD exciton chirality method.

The plant material of *C. multistachys* was collected from Songyang County of Zhejiang Province of China. Air-dried herb powder (1.5 kg) was extracted with 95% EtOH three times, and the crude was then partitioned between EtOAc and H₂O to give an EtOAc-soluble fraction, which was further defatted over a column of MCI gel eluted with 80% methanol in water. Repeated chromatography of



Multistalide A (**1**),⁷ a white amorphous powder, had a molecular formula $C_{33}H_{38}O_{10}$ as determined by the HREIMS ion at 594.2470 [M]⁺ (calcd 594.2465) with 15 degrees of unsaturation, which was supported by the positive mode of ESIMS at *m*/*z* 617.4 [M+Na]⁺ and 1211.5 [2M+Na]⁺, and the negative mode of ESIMS at 593.5 [M–H]⁻. The IR spectrum exhibited absorptions for hydroxy (3437 cm⁻¹) and carbonyl (1728 cm⁻¹) groups. The ¹H NMR spectrum of **1** (Table 1) showed the presence of three tertiary methyls (δ 0.84, 0.97, 2.08, each 3H, s), one methoxy (δ 3.80, 3H, s), and one terminal double bond (δ 5.43, 6.22, each 1H, s). In accord with the molecular formula, the ¹³C NMR spectrum (Table 1) showed 33 carbon resonances, which were further categorized by DEPT experiments into four carbonyls, four methyls, two persubstituted double bonds, one terminal double bond, six sp³ methyl-









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Table 1				
NMR data for multistalides A	and	В	(1-2) in	CDCI

	1		2		
	$\delta_{\rm H}$, mult. (J in Hz)	δ_{C}	$\delta_{\rm H}$, mult. (J in Hz)	δ_{C}	
1	2.04 (m)	26.3	2.08 (m)	25.9	
2α	0.95 (m)	10.0	0.99 (m)	16.0	
2β	1.23 (m)		0.32 (m)		
3	1.86 (m)	30.7	1.83 (m)	24.8	
4		78.0		142.5	
5		160.4		132.0	
6		124.8	3.92 (s)	40.9	
7	4.13 (s)	49.0		130.9	
8		207.9		200.1	
9	4.25 (s)	80.3	3.94 (s)	80.0	
10		49.1		51.1	
11		136.3		147.6	
12		167.6		171.0	
13a	5.43 (s)	126.7	1.94 (s, 3H)	20.5	
13b	6.22 (s)				
14	0.97 (s, 3H)	20.5	1.01 (s, 3H)	15.4	
15α	1.86 (m)	40.4	2.81 (dd, 16.4, 1.3)	25.2	
15β	2.74 (dd, 13.1, 6.9)		2.58 (m)		
1′	1.49 (m)	25.4	1.62 (m)	25.5	
2′α	0.76 (m)	16.3	0.72 (m)	11.7	
2′β	0.68 (m)		1.27 (m)		
3′	1.27 (m)	22.5	1.47 (m)	28.2	
4′	1.61 (m)	45.2		77.3	
5′	2.00 (m)	52.8	1.86 (m)	60.0	
6'α	2.78 (dd, 18.1, 6.4)	26.3	2.32 (dd, 18.3, 6.1)	22.3	
6 ′β	2.63 (dd, 18.1, 12.9)		2.70 (dd, 18.3, 13.5)		
7′		164.4		168.2	
8′		87.6		93.4	
9′	2.63 (dd, 10.7, 6.9)	49.3	1.92 (m)	55.0	
10′		44.2		44.8	
11′		129.1		127.2	
12'		171.0		172.4	
13'a	4.22 (d, 13.8)	55.3	4.42 (d, 13.5)	54.9	
13′b	4.18 (d, 13.8)		4.34 (d, 13.5)		
14′	0.84 (s, 3H)	22.7	0.88 (s, 3H)	26.2	
15'a	4.25 (d, 10.9)	65.6	4.08 (d, 11.6)	71.5	
15′b	4.20 (d, 10.9)		3.82 (d, 11.6)		
a	0.00 / 0VV	171.5	0 40 4 OV	171.3	
b	2.08 (s, 3H)	20.8	2.13 (s, 3H)	20.8	
OMe	3.80 (s, 3H)	52.8	3.80 (s, 3H)	52.8	

enes, nine sp³ methines, and four sp³ quaternary carbons. The above-mentioned functionalities accounted for seven degrees of unsaturation, and the remaining eight degrees of unsaturation required **1** being octacyclic. Two sets of spin systems being assignable to the presence of 1,2-disubstituted cyclopropane rings were evident from the ¹H NMR spectrum (H-1: $\delta_{\rm H}$ 2.04; H-2 α : $\delta_{\rm H}$ 0.95; H-2 β : $\delta_{\rm H}$ 1.23; H-3: $\delta_{\rm H}$ 1.86 and H-1': $\delta_{\rm H}$ 1.49; H-2' α : $\delta_{\rm H}$ 0.76; H-2' β : $\delta_{\rm H}$ 0.68; H-3': $\delta_{\rm H}$ 1.27). The aforementioned analysis and the fact of the existence of sesquiterpenoid dimers within this plant genus⁵ suggested that **1** is a lindenane-type sesquiterpenoid dimer.

The extensive analysis of 1D and 2D NMR (including ¹H–¹H COSY, HSQC, HMBC) spectral data allowed the assignment of com-



Figure 2. ${}^{1}H{-}^{1}H$ COSY (\longrightarrow) and selected HMBCs ($H{\rightarrow}C$) of multistalide A (1).



Figure 3. Key ROESY correlations $(H \leftrightarrow H)$ of multistalide A (1).

pound **1**. After the assignment of all protons to their directly bonded carbon atoms via a HSQC experiment, four proton-bearing spin coupling units of the molecule as drawn with bold bonds (Fig. 2) were established by ${}^{1}H{-}^{1}H$ COSY spectrum. These four structural subunits and most of other functional groups were then connected by HMBC spectrum (Fig. 2), in which the HMBCs from H₂-13 to C-12, C-11, C-8, and C-7 suggested the presence of a $\Delta^{11(13)}$ terminal double bond. The methoxyl group was placed at C-12 by the HMBC between C-12 and OMe. The presence of an α , β -unsaturated γ -lactone formed between C-8' and C-12' via an oxygen atom was evident from the down-field shifted C-8' at $\delta_{\rm C}$ 87.6 as in the case of chloramultilide A.^{5b} The only acetyl group was located at C-15' by the HMBC between H₂-15' and the carbonyl of acetyl. The remaining three oxygenated sp³ carbons were assigned to C-4 at $\delta_{\rm C}$ 78.0, C-9 at $\delta_{\rm C}$ 80.3, and C-13' at $\delta_{\rm C}$ 55.3 bearing hydroxyl groups, respectively, by the multiple HMBCs.

The relative configuration of **1** was established by the performance of a ROESY experiment (Fig. 3), in which the ROESY correlations of H-1/H-3, H-1/H-2 α , H-3/H-2 α , H-1/H-9, H-1'/H-3', H-1'/H-2' α , H-3'/H-2' α , and H-3'/H-5' indicated that H-1, H-3, H-9, H-1', H-3', and H-5' were co-facial and randomly assigned in an α -configuration. In consequence, the ROESY correlations of Me-14/H-2 β , H-9'/Me-14', H-4'/Me-14', Me-14'/H-2' β , H-2' β /H-4', H-13a/Me-14, and H-7/H-5' revealed that Me-14, Me-14', H-4', H-9', and the C-7–C-11 bond were β -oriented. A β -direction for the C-8'–O bond was tentatively assigned on the biogenetic reasoning that compound **1** was formed via an enzymatic Diels–Alder cycloaddition of two molecular lindenane-type sesquiterpenoids.^{5e,8}

Multistalide B (2)⁹ was obtained as a white amorphous powder and possessed a molecular formula C₃₃H₃₈O₁₀ as established by HRESIMS ion at 617.2379 [M+Na]⁺ (calcd for C₃₃H₃₈O₁₀Na



Figure 4. Selected HMBCs $(H \rightarrow C)$ of multistalide B (2).



Figure 5. Key ROESY correlations $(H \leftrightarrow H)$ of multistalide B (2).



Figure 6. CD and UV spectra of multistalides A and B (1-2) measured in MeOH, and the stereoview of 1, arrows denote the electric transition dipole of the chromophores.

617.2363). The IR spectrum exhibited absorptions of hydroxyl (3438 cm^{-1}) and carbonyl (1736 cm^{-1}) groups. The ¹H and ¹³C NMR data of 2 (Table 1) showed many similarities to those of shizukaol D,^{5c} except for the down-field-shifted proton (H-3': $\Delta \delta_{\rm H}$ 0.37; H-2' β : $\Delta \delta_{\rm H}$ 0.44; Me-14': $\Delta \delta_{\rm H}$ 0.22) and carbon (C-4': $\Delta \delta_{\rm C}$ 34.3) resonances scattered around C-4' as compared with those of shizukaol D,^{5c} suggesting that a hydroxyl group was attached to the C-4' of compound 2. This assignment was further confirmed by 2D NMR spectra, especially HMBC spectrum (Fig. 4), in which the quaternary C-4' resonated at $\delta_{\rm C}$ 77.3 showed HMBCs with H-5', H-2' α , and H₂-15'. The relative configuration of **2** was fixed by a ROESY experiment (Fig. 5), in which the 4'-OH was assigned as a β -configuration by comparing the chemical shift of C-4' ($\delta_{\rm C}$ 77.3) of **2** with that of chloramultilide A^{5b} bearing a β -OH at C-4' (δ_{C} 76.9). The presence of a 4 β -OH in **2** was confirmed by the ROESY correlations of H_2 -15'/H-3' and H_2 -15'/H-5'.

The absolute stereochemistry of multistalide A (1) was determined on the basis of CD spectrum (Fig. 6). The Cotton effects (206 and 234 nm) centered at 221 nm corresponding to the UV maximum of α,β -unsaturated γ -lactone (C-7', C-11', and C-12', Woodward's rules showed ca. 227 nm).¹⁰ The first negative Cotton effect at 234 nm and the second positive Cotton effect at 206 nm arising from the exciton coupling between the α,β -unsaturated γ -lactone and the α,β -unsaturated ester group (C-11, C-12, and C-13, Woodward's rules showed ca. 203 nm)¹⁰ indicated a negative chirality for **1**. The absolute configuration of **1** was thus established as depicted, which is consistent with those of chlorahololides A and B^{5e} isolated from this genus.

The CD curve of multistalide B (2) was very similar to those of the known sesquiterpenoid dimers, chlorahololides A and B,^{5e} in the range of 210-265 nm (Fig. 6). The first positive Cotton effect at 257 nm and the second negative Cotton effect at 207 nm showed a positive chirality for compound 2. The absolute configuration of 2 was thus defined as depicted.

Acknowledgments

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Supplementary data

Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.tetlet.2009.12.001.

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- Multistalide A (1): white, amorphous powder; $[\alpha]_D^{20}$ –124 (*c* 0.370, CHCl₃); UV (MeOH) λ_{max} (log ε) 221 (4.26) nm; CD (MeOH) λ_{max} ($\Delta \varepsilon$) 292 (-3.78), 264 (-1.14), 234 (-7.37), 206 (4.63) nm; IR (KBr, disc) λ_{max} 3437, 2922, 1728, 1630, 1441, 1383, 1238, 1113 cm⁻¹; ¹H and ¹³C NMR see Table 1; EIMS 70 eV m/z 594 [M]⁺ (51), 577 (65), 499 (100), 403 (88), 105 (86); HREIMS *m*/*z* 594.2470 [M]⁺ (calcd for C₃₃H₃₈O₁₀, 594.2465). Oikawa, H.; Suzuki, Y.; Naya, A.; Katayama, K.; Ichihara, A. *J. Am. Chem. Soc.*
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- Multistalide B (**2**): white, amorphous powder; $[\alpha]_D^{20} -113$ (*c* 0.155, CHCl₃); UV (MeOH) λ_{max} (log ε) 218 (4.08) nm; CD (MeOH) λ_{max} ($\Delta \varepsilon$) 257 (21.8), 207 (-43.2) nm; IR (KBr, disc) λ_{max} 3438, 2924, 1736, 1436, 1379, 1238, 1086 cm⁻¹; IL and ¹³C NMB cco Table 1. FCl MeCl (1.11) (¹H and ¹³C NMR see Table 1; ESI MS (positive) m/z 617.4 [M+Na]⁺, 1211.5 [2M+Na]⁺; HRESIMS *m*/*z* 617.2379 [M+Na]⁺ (calcd for C₃₃H₃₈O₁₀Na, 617.2363).
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